

## Characterisation of Subvisible Particles and Nanomaterials with Nanoparticle Tracking Analysis and Dynamic Light Scattering



Dr. Markus Epe, Field Application Scientist EURA

**Dr. Diogo Fernandes, Product Technical Specialist** 

### Overview



Introduction into Nanoparticle Tracking Analysis

**Dynamic Light Scattering** 

Application Example NTA/DLS

# Nanosight

# nanoparticle tracking analysis



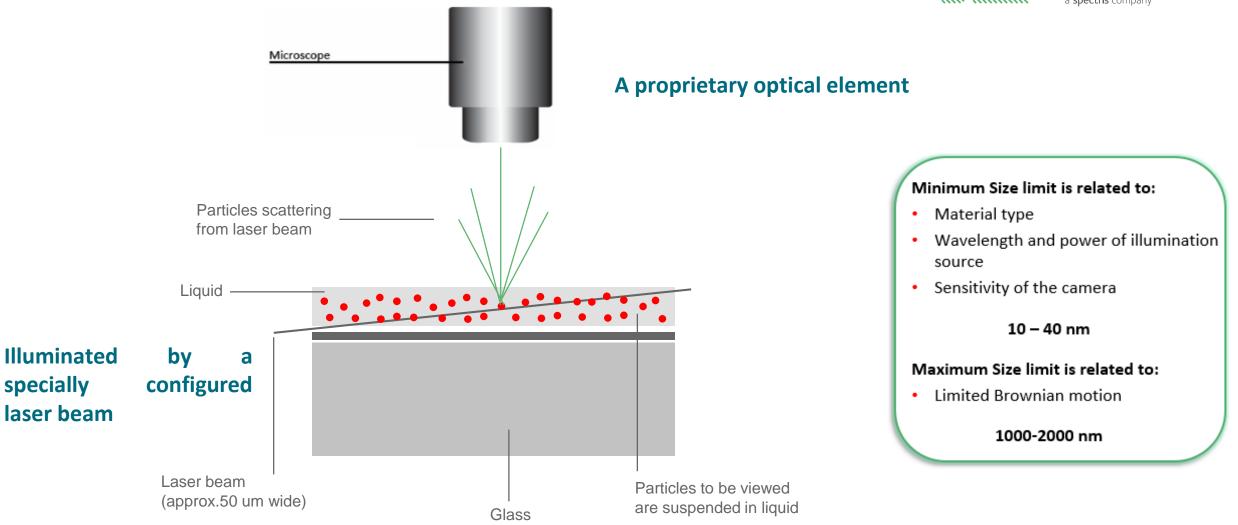


## Principle and Set up



# NANOSIGHT TECHNOLOGY



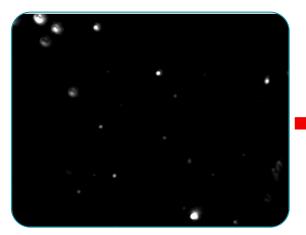


Schematic of laser sample chamber

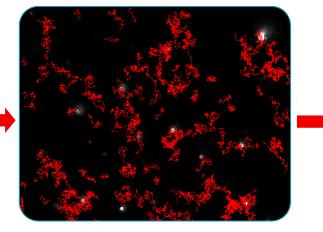
## NTA Experimental Protocol

1. Capture: The software captures a movie file of the particles moving produce of the particle

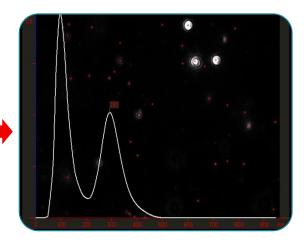
 Tracking: The software locates each particle and tracks the motion of each particle independently, yielding mean square displacement.
Analysis: Application of the Stokes Einstein equation converts mean square displacement to particle size. The distribution is an accumulation of all the single particle measurements.



Capture (~60 sec)



Tracking



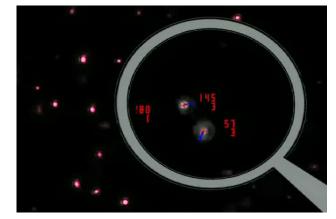
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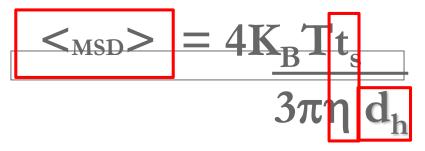
Analysis

## Sizing: Stokes-Einstein

- Measure particles' mean square displacement (MSD) due to Brownian motion
- Calculated parameter is particle's sphere equivalent hydrodynamic diameter.
- Temperature measured and appropriate viscosity used.
- Absolute method no calibration required
- Independent of refractive index, density or mass.

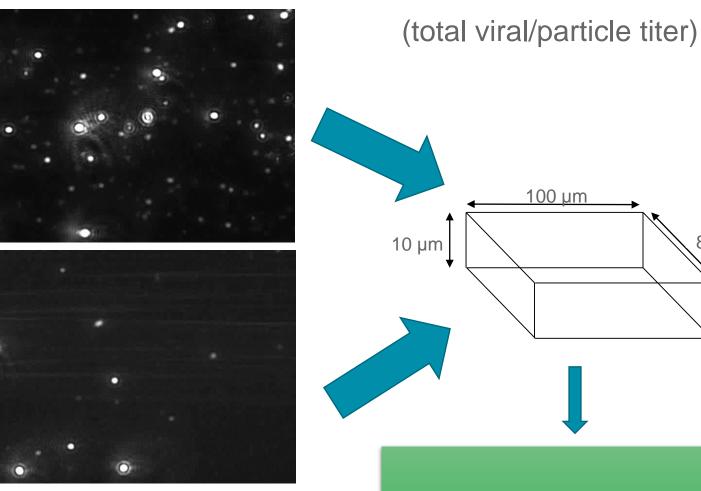






Stokes-Einstein equation  $K_B$  = Boltzmann Constant T = temperature  $t_s$  = sampling time  $\eta$ = viscosity  $d_h$  = hydrodynamic diameter

## Concentration: Quantify particles in known volume



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**Absolute number concentration** 

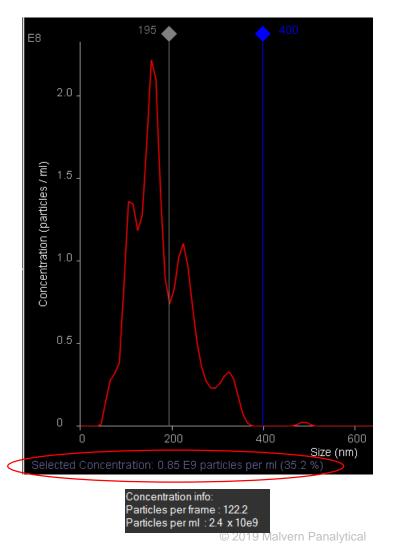
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80 µm

## Concentration of selected range

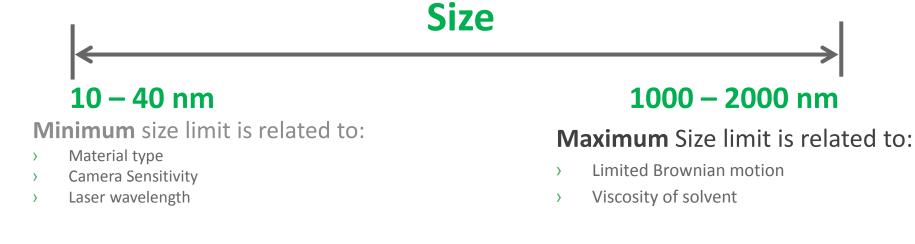


- Concentration can be reported for any subset
- Data is accumulation of individual particle measurements, so any statistical measure can be applied.



## NTA Specifications: Size and Concentration Limits





## Concentration

~ 10<sup>6</sup>-10<sup>7</sup> particles/ mL

Minimum is related to:

> Poor statistics (Requiring longer analysis time)

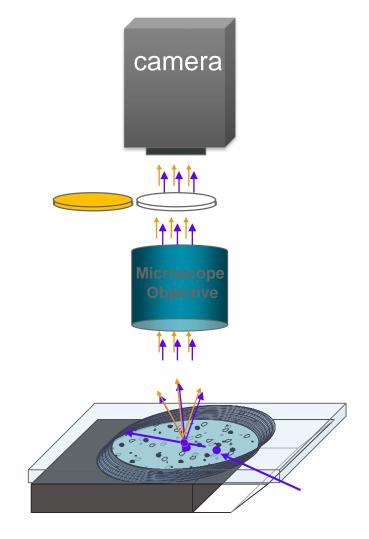
#### ~ 10<sup>9</sup> particles/ mL

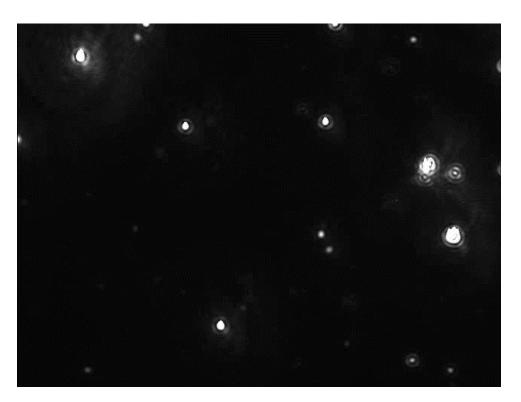
Maximum is related to:

- > Inability to resolve neighboring particles
- > Tracks too short before crossing occurs

## Fluorescence measurement (optional)



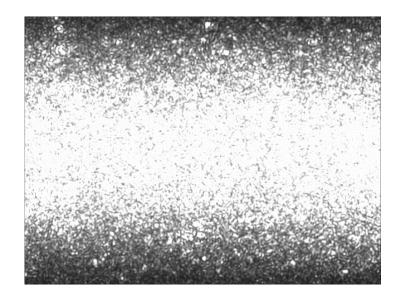




#### Video of fluorescence process

## Analysis in Complex Biological Media

100nm fluorescent particles in FBS



# Scatter Mode (all particles)

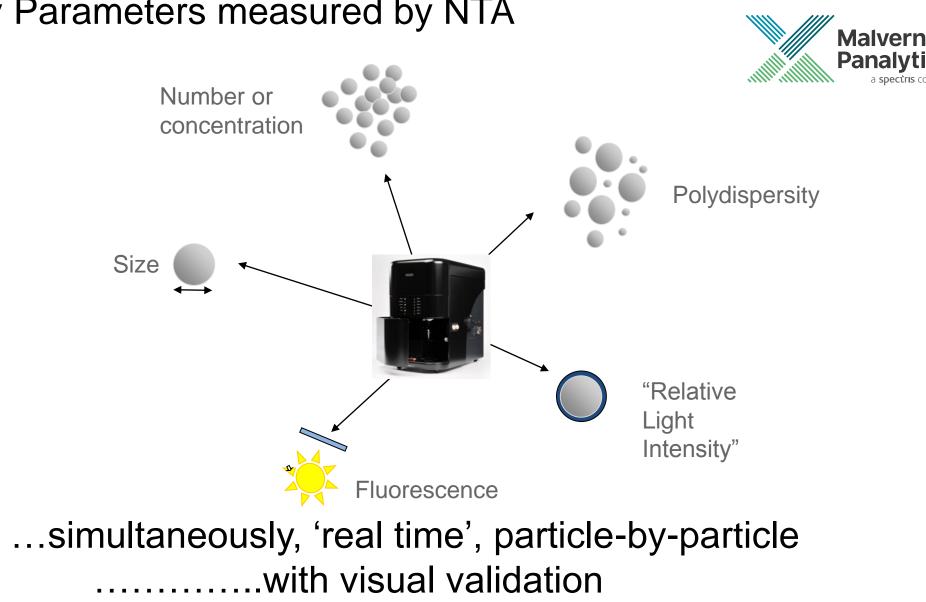


#### Fluorescence Mode (only labelled particles)





Available laser sources (nm)	Standard filter provided (nm)	Fluorophores known to work based on customer feedback	
Violet 405	430 long pass	-QD605 -QD625 -QD705	
Blue 488	500 long pass	-AlexaFluor 488 -PE -E-GFP / GFP -DiO -FITC	-PerCP -FM1-43 -Anti-HA (H5N1) -PKH67
Green 532	565 long pass	-PE -Nile-Red -Spiro-Red -Cy3	-Rhodamine-PE -Alexa 546 -Dil -PKH26
Red 638	650 long pass	-AlexaFluor 647	



#### Summary Parameters measured by NTA

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# News: Zetasizer Ultra advanced features



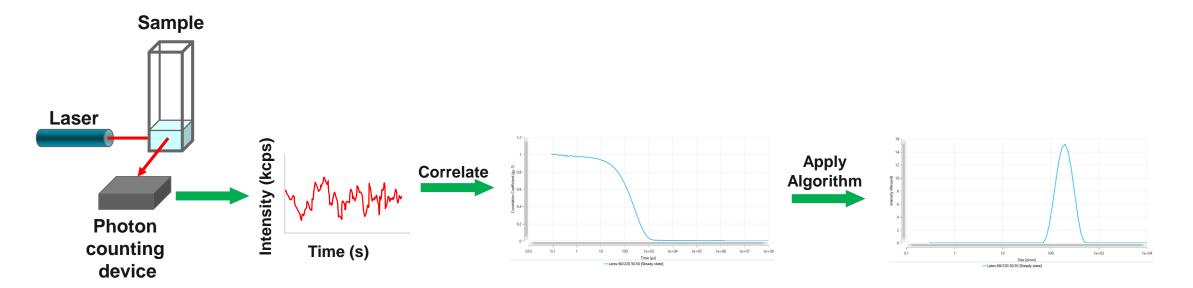
## **Dynamic Light Scattering**

#### What is it and How does it work?



- Non-invasive technique for measuring the size of particles and molecules dispersions
- Analyses the time-dependence in the intensity of the scattered light (auto correlation) to determine their diffusion speed (Brownian motion) and subsequently their hydrodynamic size

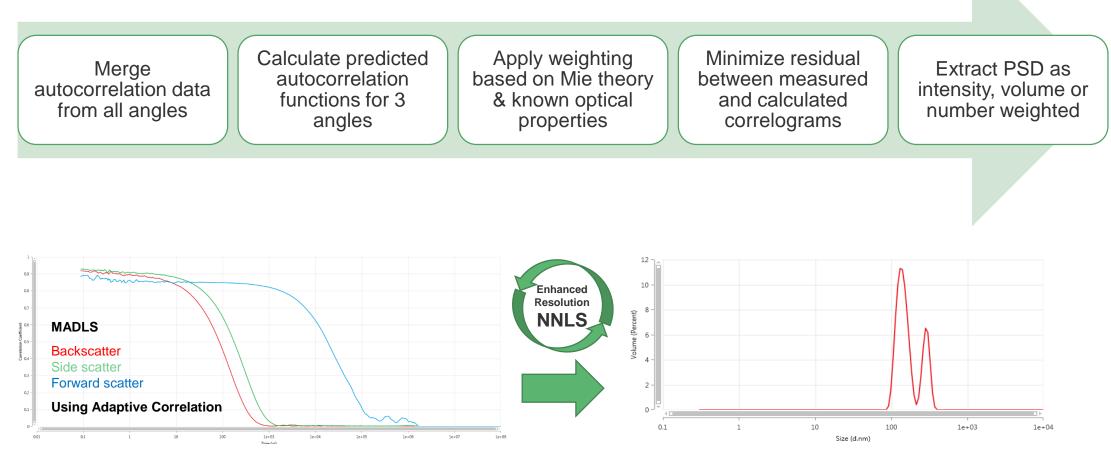
(A. Einstein (1926) Investigations on the theory of the Brownian movement. In: Fürth R., ed. Cowper, A.D., translator. Methuen, London, 124)



# Multi Angle Dynamic Light Scattering



#### How it Works



## **Particle Concentration**



- Particle concentration measurements are an extension of MADLS, and require no calibration
- The total scattering detected from the sample is converted into number of particles per mL by:
  - Using the PSD obtained from MADLS to determine how much each individual population is scattering
  - Using the PSD to calculate the scattering cross-section and amount of scattering per particle
- Instrument detection efficiency is normalized using toluene (known Rayleigh ratio)

Complementary technologies:

Zetasizer Ultra and NTA



# Comparing DLS and NTA



- Understanding what each instrument does well, what are their limitations is the basis of deciding which instrument to select and when both may provide additional information.
  - > DLS Strengths:
    - Dynamic Size Range
    - Concentration Dynamic Range (NIBS)
    - Ease of Use
    - Routine Zeta potential measurement
    - Recognized technology.
    - Very good trend spotter. Microrheology

- > NTA Strengths:
  - Higher resolving power (peakto-peak) due to single particle measurement.
  - Measurement of particle concentration.
  - Fluorescence mode speciation
  - Less bias towards aggregates in sample.
  - Image of particles provides valuable information.
  - Measures particle diffusion coefficient.

## Why use both?



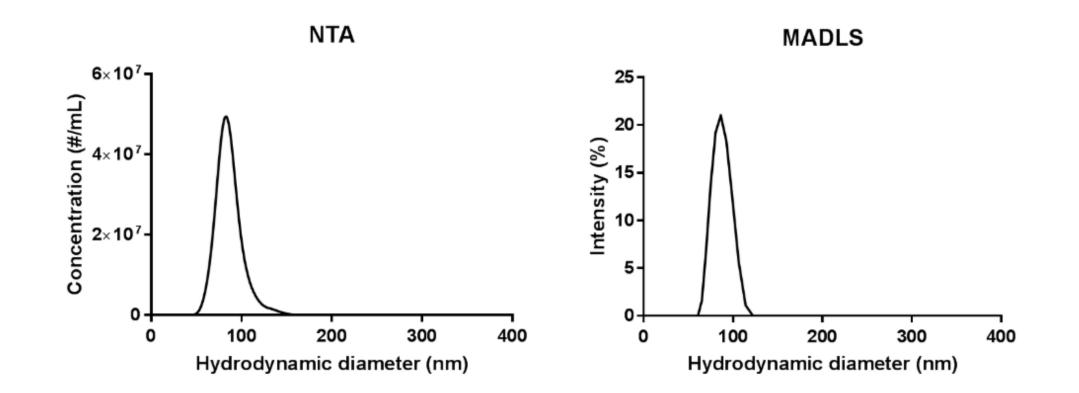
- Limitations of one addressed by the other
- Complementary data
- How can I trust my data?
- Is the data affected in any way by the measurement technology?
- Or by the user; through sample preparation or analysis parameters employed?
- Together = Comprehensive suite of measurement parameters





#### Particle Distribution DLS and NTA 1:10000 dilution Liposomes

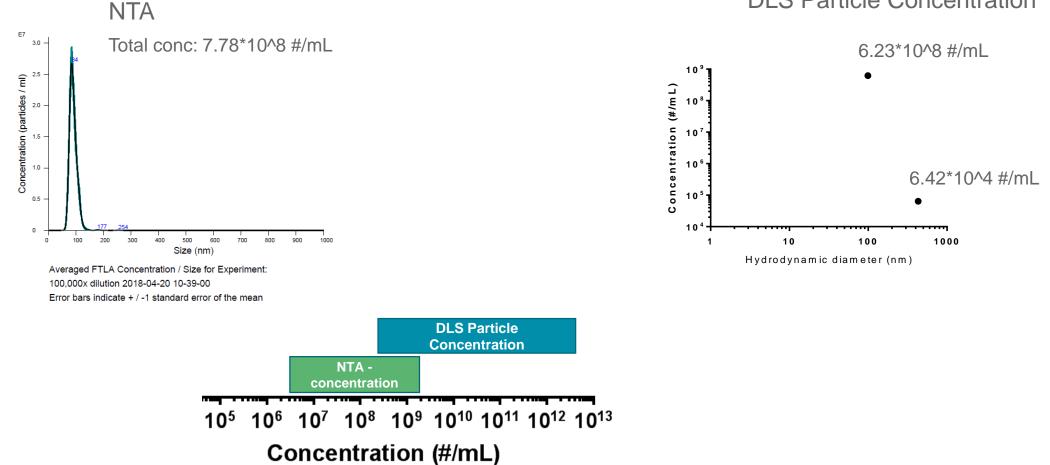




#### **Particle Concentration** Liposomes (36.5mg/mL HSPC/CHOL 55:45 Liposomes) down to 1:10,000,000

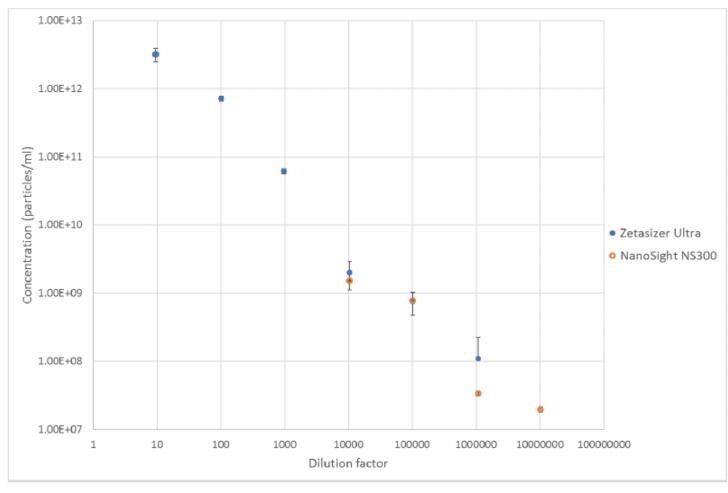


#### **DLS Particle Concentration**



## Particle Concentration

#### Liposomes measured with NTA and DLS





#### Measured with DLS and NTA

Figure 3 Concentration results from measurements of liposomes on a Zetasizer Ultra and Nanosight NS300.

### Application Note AN180519

May 27, 2019

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<u>markus.epe@malvern.com</u> <u>diogo.fernandes@malvernpanalytical.com</u> www.malverpanalytical.com

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**Zit**le of the presentation

May 27, 2019